

leaves suggests the possible disproportionation of one to the other noted earlier<sup>10</sup> for *P. balsamifera* bark. It is interesting to note that *P. balsamifera* leaves contained no salireposide whatsoever.

### EXPERIMENTAL

**Materials.** Fresh leaves from a *P. balsamifera* tree cut in Langlade County, Wisconsin on June 17, 1968 were processed within a few hours with EtOH by the Waring Blendor procedure. *P. trichocarpa* leaves were stripped from trees in Pierce County, Washington on June 13, 1969, placed immediately into 95% EtOH, and processed a few days later by the Waring Blendor procedure.

**Isolation and identification of components.** The following crystalline components were isolated from the eluate fractions and identified by mixed m.p. and identity of IR spectra with authentic material indicated by reference: salicin<sup>11</sup>, 1-*O*-*p*-coumaroyl-glucose,<sup>12</sup> tremuloidin,<sup>11</sup> trichocarpin,<sup>13</sup> tremulaein,<sup>14</sup> salireposide,<sup>15</sup>  $\omega$ -salicyloylsalicin,<sup>16</sup> *d*-catechin,<sup>6</sup> and isoquercitrin.<sup>6</sup>

**Chromatography of flavonoid components.** Flavonoid components of eluate fractions were monitored by means of TLC on Polygram Cell developed with 30% AcOH and sprayed with ethanolic KOH. Plates were examined under visible and UV light before and after spraying.

The flavonoid glucosides were separated preparatively on Whatman 3M paper developed with the upper layer of 4:1:5 *n*-BuOH-AcOH-H<sub>2</sub>O at 25°.

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<sup>10</sup> I. A. PEARL, *Tappi* **52**, No. 3, 428 (1969).

<sup>11</sup> I. A. PEARL and S. F. DARLING, *J. Org. Chem.* **24**, 731 (1959).

<sup>12</sup> I. A. PEARL and S. F. DARLING, *Tappi* **50**, No. 7, 324 (1967).

<sup>13</sup> I. A. PEARL and S. F. DARLING, *Phytochem.* **7**, 1951 (1968).

<sup>14</sup> I. A. PEARL and S. F. DARLING, *Phytochem.* **10**, 483 (1971).

<sup>15</sup> I. A. PEARL and S. F. DARLING, *J. Org. Chem.* **24**, 1616 (1959).

<sup>16</sup> I. A. PEARL and S. F. DARLING, *Arch. Biochem. Biophys.* **102**, 33 (1963).

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## SAPINDACEAE

### CHEMICAL INVESTIGATION ON THE LEAVES OF *EUPHORIA LONGANA*

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*Plant.* *Euphoria longana* Lam. (Syn. *Nephelium longana* Cambess., Longan). Sapindaceae.

*Uses.* Medicinal.<sup>1</sup>

*Previous work.* On seeds,<sup>2</sup> stem and leaves.<sup>3</sup>

*Leaves.* Extracted with light petroleum, chloroform and ethanol.

*Petroleum ether extract.* This was chromatographed on alumina. Petroleum fraction on further purification by chromatography and crystallization afforded friedelin,<sup>4</sup> C<sub>30</sub>H<sub>50</sub>O

<sup>1</sup> *The Wealth of India*, Volume III, p. 230, C.S.I.R., New Delhi (1952).

<sup>2</sup> J. GEDEON and F. A. KINCL, *Arch. Pharm.* **289**, 162 (1956).

<sup>3</sup> T. TSUKAMOTO, T. TOMINAGA and J. TAKAHASHI, *J. Pharm. Soc.* **69**, 40 (1949).

<sup>4</sup> E. J. COREY and J. J. URSPRUNG, *J. Am. Chem. Soc.* **78**, 5041 (1956).

(m.p., mixed m.p.  $[\alpha]_D$ , IR, TLC, oxime). Petroleum–benzene (1:1) fraction on rechromatography yielded 16-hentriacontanol,  $C_{31}H_{64}O$  (m.p., mixed m.p.) and epifriedelinol,<sup>5</sup>  $C_{30}H_{52}O$  (m.p., mixed m.p.  $[\alpha]_D$ , IR, m.p. and  $[\alpha]_D$  of acetate). Benzene and benzene–ether (1:1) fractions yielded  $\beta$ -sitosterol,<sup>6</sup>  $C_{29}H_{50}O$  (m.p., mixed m.p. and  $[\alpha]_D$ ; m.p., mixed m.p. and  $[\alpha]_D$  of acetate) and stigmaterol,<sup>6</sup>  $C_{29}H_{48}O$  (m.p., mixed m.p. and  $[\alpha]_D$ ; m.p., mixed m.p. and  $[\alpha]_D$  of acetate).

*Ethanol extract.* The ethanolic extract concentrated, pulped, dried and exhaustively extracted with  $Et_2O$ . The  $Et_2O$  extract concentrated and kept in a refrigerator when greenish precipitate accumulated in the flask. The precipitate on repeated crystallization (decolourizing carbon) afforded stigmasteryl D-glucoside,<sup>7</sup> m.p. 290–293 (decomp.) (Liebermann–Burchard and Molisch test positive). The glucoside on hydrolysis with 4%  $H_2SO_4$  yielded stigmaterol (m.p. and mixed m.p.) and D-glucose (identified by paper chromatography).

<sup>5</sup> V. ANJANEYULU, D. N. RAO and L. R. ROW, *J. Indian Chem. Soc.* **44**, 123 (1967).

<sup>6</sup> L. F. FIESER and M. FIESER, *Steroids* p. 351, Reinhold, New York (1959).

<sup>7</sup> A. NITTA, *Yakugaku zasshi* **85**, (2), 173 (1965) (Japan); *Chem. Abs.* **62**, 14615 b (1965).

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## SCROPHULARIACEAE

### AN UNUSUAL ANTHOCYANIN IN *ANTIRRHINUM MAJUS*

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(Received 9 June 1970, in revised form 8 January 1971)

**Abstract**—In three genetic stocks of *A. majus*, cyanidin-3-glucoside has been found to occur along with the normal pigment, cyanidin-3-rutinoside.

THE ANTHOCYANINS of the garden snapdragon hitherto investigated are of two types, either cyanidin-3-rutinoside (antirrhinin) in the normal form (magenta or crimson flowers) or pelargonidin-3-rutinoside in the recessive mutant *eosinea* (pink or bronze flowers).<sup>1,2</sup> Other pigments, such as aurones and flavones, also contribute to the final flower colour, so that a large range of shades is possible.

## RESULTS

Flowers from three lines (viz. an inbred derivative of the commercial variety 'Eclipse'; 'Black Prince'; and an inbred derivative of 'Pan Crimson') were investigated using TLC on cellulose. All three were found to contain a second anthocyanin which was purified by

<sup>1</sup> R. SCOTT-MONCRIEFF, *Biochem. J.* **24**, 753 (1930).

<sup>2</sup> R. SCOTT-MONCRIEFF, *J. Genet.* **32**, 117 (1936).